Different bat detectors and processing software... Same results?

Santiago Perea¹*, Elena Tena¹

¹Departamento de Biodiversidad, Ecología y Evolución. Facultad de Ciencias Biológicas. Universidad Complutense de Madrid.

*Corresponding author: santip1320@gmail.com

DOI: https://doi.org/10.14709/BarbJ.13.1.2020.01

Keywords: acoustic, Chiroptera, detection, echolocation, noise filtering

ABSTRACT

There has been an increase in commercial bat detectors and noise filtering software for monitoring bat activity. In this study, we compare the recording efficiency of three bat detectors from the popular brand Wildlife Acoustics (Echo Meter 3, Echo Meter Touch Pro 1 and Song Meter 2 BAT) and the effectiveness of two noise filtering software (Kaleidoscope and SonoBat Batch Scrubber). To do so, we recorded 7513 files from 13 urban parks in Madrid in 2017, that were manually identified to species level. The results show that the Echo Meter 3 records significantly less activity than the Echo Meter Touch Pro 1 and Song Meter 2 BAT. Our results also identify SonoBat Batch Scrubber as more reliable than Kaleidoscope for preventing false negatives. Therefore, our study demonstrates that different bat detectors, and different noise filtering software, can provide different results.

INTRODUCTION

Monitoring bats echolocation using bat detectors is a common, non-intrusive technique for studying the behaviour and ecology of bats (Collins & Jones 2009). This can be used to complement other techniques such as roost surveys or direct capture (Flaquer et al. 2007, Fenton 2013). It facilitates the collection of large quantities of data which is used by researchers to determine the presence of bats, species identification, to investigate their behaviour and differences in habitat use, as well as estimating relative abundance or quantifying activity (Adams et al. 2012, Russo & Voigt 2016).

Nevertheless, sampling bats using bat detectors has a series of limitations that must be considered. Variability amongst sampling methodologies, detector models, microphones and detection algorithms can significantly affect the detection efficiency among bat detectors, as demonstrated by Adams et al. (2012) between bat detector brands. In addition, the echolocation calls produced by bats is highly variable between species which can result in interspecific differences in detection efficiency (Adams et al. 2012, Russo & Voigt 2016). Other sources of variability include intraspecific variation in call structure, which is influenced by factors such as habitat, the presence of conspecifics, environmental noise, age, sex, geographic variation or existence of sympatric acoustic variants (Walters et al. 2012, Barataud 2015, Russo et al. 2018).

Since the late 1990s, the acoustic identification of bat species has undergone a radical change thanks to assisted identification systems and noise filtering software (Russo & Voigt 2016). These systems are based on algorithms that help classify recordings (e.g. as noise or to species) to reduce the demand for manual processing. These changes have promoted an increased use of bat detectors and encouraged subsequent improvements to their recording capacity. Nowadays, there are two types of bat detectors commonly used: hand-held bat detectors, for active use such as walking transects and roost surveys; and static bat detectors, that are now capable of recording continuously for many hours generating large acoustic datasets, which in turn feed the increased need for these kinds of assisted identification and noise filtering software (Lemen et al. 2015, Rydell et al. 2017).

In this study we have two aims: (i) to compare the number of bat recordings captured between three models of Wildlife Acoustic bat detectors (Wildlife Acoustics Inc., U.S.A.; Echo Meter 3, Echo Meter Touch Pro 1, Song Meter Bat 2); and (ii) to compare the efficiency and accuracy of two noise filtering software (Kaleidoscope 4.5.4 (Wildlife Acoustics Inc., U.S.A.) and SonoBat Batch Scrubber 5.1 (SonoBat. Bat Call Analysis Software). Both the bat detectors and noise filtering software vary in price and release date.

MATERIAL AND METHODS

Study site

Sampling was conducted across 13 urban parks in Madrid, Spain (Fig. 1) during July 2017. One sampling period was carried out once per park and consisted of multiple ten-minute sampling stations in order to determine the presence and activity of bats per park. The number of sampling stations per park was determined by the park size (Tena et al. 2019) (see Supplementary Table 1 for sampling stations per park and park size). All bat detectors used were positioned...
to record simultaneously at each sampling station, with the same orientation and inclination to avoid biases.

**Equipment and recorder conditions**

Three Wildlife Acoustics bat detectors were used in this study; two hand-held bat detectors (Echo Meter 3 and Echo Meter Touch 1 Pro) and one static bat detector (Song Meter 2 BAT) with a SMX-US ultrasonic microphone (see Table 1 for bat detector settings).

**Manual call processing**

For the manual identification, each recording was split into five second sequences, using Kaleidoscope 4.5.4 (Wildlife Acoustics Inc., U.S.A.), in order to standardize the bat call study (Bertran et al. 2019). A bat pass consisted of at least two echolocation pulses above -60 dB (Russ 2012). Audacity 2.2.1 was then used to analyse the recordings. The following parameters were measured manually (Rydell et al. 2017) from each pulse to identify species (Russo & Jones 2002, Barataud 2015): start frequency, end frequency, frequency of maximum energy, duration and inter-pulse interval. Recordings were discarded where it was not possible to determine the species identification. In this study, noise was considered as recordings with no echolocation pulses or less than 2 pulses above -60dB. The total number of bat passes and noise were summarized for each bat detector.

**Noise filtering software**

Two types of commercial software were selected to compare the effectiveness of noise filtering: a free license software, Kaleidoscope 4.5.4, and a paid license software, SonoBat Batch Scrubber 5.1 (SonoBat. Bat Call Analysis Software). For Kaleidoscope, the signal parameters were configured as follows: 8-120 kHz, 2-500 ms, 500 maximum inter-syllable gaps and 2 minimum number of pulses. For SonoBat Batch Scrubber the default settings were selected: medium, accepts all calls except low quality calls; accepts some noise with tonal content and includes from 5-20 kHz signals.

**Statistical procedures**

A heterogeneity test was performed to compare the total number of pulses recorded by each bat detector. Non-parametric Chi-square test were performed to compare the total number of bat passes between bat detectors and the total number of bat passes by species for each bat detector. Non-parametric Chi-square test were also performed to compare the correct and incorrect ID bat and ID noise between both software and manual identification; and between noise filtering software. Statistical analysis was conducted in R 3.3.2 (R CoreTeam 2012).

**RESULTS**

In total 18 hours of recordings were collected which comprised 7513 five-second files. Of these, 2828 were manually identified as bats and 4685 as noise. The total number of echolocation pulses observed were 198688. Across the study, four different species were identified manually: Common pipistrelle (Pipistrellus pipistrellus), Soprano pipistrelle (Pipistrellus pipistrellus), Kuhl’s pipistrelle (Pipistrellus kuhlii) and Savi’s pipistrelle (Hypsugo savii).
Comparison among bat detectors

The heterogeneity test showed a significant difference in the total number of echolocation pulses recorded between bat detectors (Fig. 2; $\chi^2 = 45203.34$, $p < 0.001$), as a greater number of echolocation pulses were recorded by the EMT and SM2BAT than the EM3.

There were significant differences in the number of bat passes between bat detectors ($\chi^2 = 470.25$, $p < 0.001$) and also for $P.$ pipistrellus (Fig. 3; $\chi^2 = 280.27$, $df = 2$, $p < 0.001$), $P.$ pygmaeus (Fig. 3; $\chi^2 = 79.10$, $df = 2$, $p < 0.001$) and $P.$ kuhlii (Fig. 3; $\chi^2 = 139.45$, $df = 2$, $p < 0.001$). In contrast, there was no significant difference for $H.$ savii (Fig. 3; $\chi^2 = 5.44$, $df = 2$, $p = 0.657$), which was the least recorded across the study.

![Fig. 2 - Total number of echolocation pulses observed for each detector.](image)

![Fig. 3 - Total number of bat passes observed by species for each detector. Pipi: Pipistrellus pipistrellus; Pipy: Pipistrellus pygmaeus; Piku: Pipistrellus kuhlii; Hysa: Hypsugo savii.](image)

Comparison between noise filtering software

There were significant differences between the number of files manually identified as a bat pass or as noise compared to those identified by both noise filtering software (Manual ID - Kaleidoscope, $\chi^2 = 184.88$, $p << 0.001$; Manual ID - SonoBat Batch Scrubber, $\chi^2 = 8.44$, $p = 0.004$). There was a significant difference in the number of files classified as bat passes and noise between the two noise filtering software ($\chi^2 = 2867.44$, $p << 0.001$). The results show Kaleidoscope has higher true negatives (correctly ID noise) but higher false negatives (incorrectly ID noise), whereas SonoBat Batch Scrubber has higher false positives (incorrectly ID bat) but lower false negatives (incorrectly ID noise) (Fig. 4).

![Fig. 4 - Comparison of correctly and incorrectly ID bat and ID noise between SonoBat Batch Scrubber and Kaleidoscope noise filtering software.](image)

DISCUSSION

Supporting previous studies (Fenton 2000, Adams et al. 2012), our results showed that recordings made by different bat detector models can vary significantly. Overall, the EM3 recorded significantly less echolocation pulses than EMT and SM2BAT, as well as bat passes for $P.$ pipistrellus, $P.$ pygmaeus and $P.$ kuhlii. It is possible no significant differences were observed for $H.$ savii due to the small sample size for this species. We also identified significant differences between manual ID and both noise filtering software and between the noise filtering software. We demonstrated that the SonoBat Batch Scrubber has a lower risk of misclassifying bat passes as noise than Kaleidoscope.

Apart from these results, there is a series of advantages and disadvantages that differ between bat detectors: SM2BAT is a static bat detector that, despite being more expensive, allows the bat detector to be programmed with a recording schedule and placed during the day, therefore eliminating the need to work at night. Hand detectors, on the other hand, are cheaper than static detectors and allow the user to observe bat activity while recording but require the user to operate them in the field.

The EMT recorded more bat passes overall, as well as for three of the four species in our study, and was not significantly different from the most expensive bat detector in our study, the SM2BAT (Fig. 3). Therefore, it provides a cost-effective alternative to the SM2BAT where it is not necessary to programme the recording schedule. Researchers using an EM3 should be aware of the risk that recorded activity is likely to be lower than true bat activity. Even though the EM3 recorded fewer bat passes than the other two bat detectors, it also recorded less noise than the EMT (see supplementary table) and there were no significant differences in the number of species recorded for this study.

Although Kaleidoscope is a free license software, we recommend researchers use SonoBat Batch Scrubber rather than Kaleidoscope as files containing bat calls is more likely to be mistakenly discarded in the latter which may result in a false representation of bat activity. However, previous studies recommend the use of more than one filtering program in order to minimize possible loss of information (Méndez et al. 2016). Different commercial, automated identification classifiers have similar variability in their effectiveness to classify bat calls to a species (Russo & Voigt 2016).
This variability inherently produces differences in results between studies. Therefore, this study supports previous work highlighting the importance of combining automated identification classifiers with manual identification to avoid possible misidentification (Russo & Voigt 2016). There are some species that due to their echolocation calls have higher probabilities of being classified as noise such as the short sweeps of Plecotus auritus flying in clutter (Rydel et al. 2017).

As our study was geographically and temporally limited, we advise future studies to conduct further studies in additional areas, over larger time periods, and between different brands and species. In doing so, these studies would highlight potential differences between other species and other recording factors.

CONCLUSIONS

In conclusion, we demonstrate that using different bat detectors, and processing software may produce different results for the same study. We recommend the SM2BAT or EMT for recording bat activity, over the EM3, as they detect greater activity. Both bat detectors offer different user requirements such as cost and programming options. For subsequent noise filtering of recording, we recommend the SonoBat Batch Scrubber rather than the license free software of Kaleidoscope to reduce the risk of false negatives.

ACKNOWLEDGEMENTS

This paper was supported by Elena Tena’s contract from the Council for Culture, Education and Sport in the Autonomous Community of Madrid and European Social Fund. We would like to thank Prof. José Luis Tellería for his ideas and suggestions, both of them so necessary. And, also, we are grateful to Dr. Óscar de Paz and the University of Alcalá for providing the materials without which this work would not have been possible. To Domingo Perea and Manuel Boado for their time in the corrections when writing the article. Finally, to the reviewers that considerably improved this paper.

REFERENCES


MÉNDEZ, D., MARTÍNEZ, S., PÉREZ G. & DE PAZ, O. (2016, December 3-4). Análisis comparativo de diferentes índices y softwares para el estudio de la quiropterofauna a través sus ultrasonidos. VI Jornadas de la Asociación Española para la Conservación y Estudio de los Murciélagos (SECEMU), Vairao, Portugal


